

DEVELOPMENT OF SPECTROPHOTOMETRICAL ASSAY FOR THE STUDY OF THE INTERACTION OF ANTIOXIDANT STANDARDS WITH 1,1-DIPHENYL-2-PICRYLHYDRAZYL FREE RADICALS

Touhami Lanez^{*}, Hadia Hemmami

*University of El Oued, VTRS Laboratory, PO Box 789, 39000,
El Oued, Algeria*

*Corresponding author: touhami-lanez@univ-eloued.dz

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Abstract: The aim of the present study was to develop a spectrophotometric based assay for the determination of binding parameters like binding constant and binding free energy of the free and 1,1-diphenyl-2-picrylhydrazyl (DPPH) bound forms of a set of antioxidant standards (AS). The determination of binding constant and binding free energy is based upon the decrease in absorbance of the electronic absorption spectrum of a 10^{-4} M acetonitrile solution of DPPH in the presence of a given concentration of a solution of antioxidant standards in the same solvent.

Keywords: *binding constants, free energy, UV-Vis*

INTRODUCTION

Scientific literature dealing with the study of the antioxidant activity of phenolic compounds and antioxidant standards, generally focuses on evaluating the antioxidant activity using different methods based on scavenging activity of free radicals like 1,1-diphenyl-2-picrylhydrazyl (DPPH) [1 – 5] and superoxide anion (O_2^-) [6 - 10]. Information on binding parameters of DPPH with antioxidant standards or with potential antioxidant compounds have never, as far as we know, been reported until now.

The aim of this work is to provide a new method, based on spectrophotometric measurements, for the determination of binding parameters of a set of four commonly used antioxidant standards with DPPH radical.

The decrease in absorbance of an acetonitrile solution of DPPH in the presence of antioxidant standards were used to investigate the interaction between DPPH and antioxidant standards.

MATERIALS AND METHODS

Chemical

Acetonitrile (ACN) (HPLC-grade from Sigma-Aldrich) was used as solvent without further purification, 1,1-diphenyl-2-picrylhydrazyl radical (DPPH) (99%), ascorbic acid (99.7 %) (AA), gallic acid (99 %) (GA), caffeic acid (99 %) (CA), rutin (97 %) (R) were all purchased from Alfa Aesar and used without further purification. All other reagents used are analytical grade.

Instrumentation and software

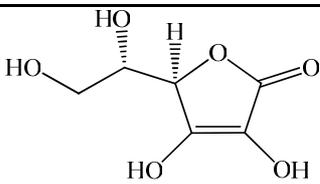
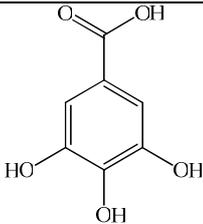
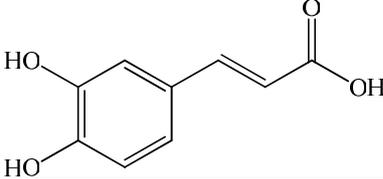
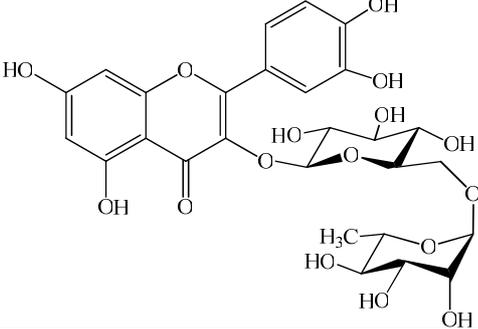
UV-Vis experiments were performed using a UV-Vis spectrometer (Shimadzu 1800) and a quartz voltammetric cell with a volumetric capacity of 5 mL. Data acquisition was accomplished with a Pentium IV (CPU 4.0 GHz and RAM 2 Gb) microcomputer using UV probe software version 2.34 (Shimadzu). Graphs plot and calculus were carried out using OriginLab software version 2.0 (Integral Software, France).

The electronic spectrum of 10^{-4} M of DPPH in acetonitrile was obtained without standard antioxidants. The spectroscopic response of the same amount of DPPH was then measured after the addition of gradually increasing concentration of a solution of the standard antioxidant in the same solvent.

Antioxidant standards

Among several commonly used antioxidant standards, four are chosen for evaluation of their binding parameters with 1,1-diphenyl-2-picrylhydrazyl free radicals (Table 1).

Table 1. Antioxidant standards (AS) used in the present study

Molecular structure	Nomenclature
	<p>Common name: Ascorbic acid IUPAC name: 1,2-dihydroxyethyl-3,4-dihydroxyfuran-2(5H)-one Formula: C₆H₈O₆</p>
	<p>Common name: Gallic acid IUPAC name: 3,4,5-trihydroxybenzoic acid Formula: C₇H₆O₅</p>
	<p>Common name: Caffeic acid IUPAC name: 3-(3,4-dihydroxyphenyl)-2-propenoic acid Formula: C₉H₈O₄</p>
	<p>Common name: Rutin IUPAC name: 2-(3,4-dihydroxyphenyl)-5,7-dihydroxy-3-[[[(2S,3R,4S,5S,6R)-3,4,5-trihydroxy-6-({[(2R,3R,4R,5R,6S)-3,4,5-trihydroxy-6-methyloxan-2-yl]oxy} methyl)oxan-2-yl]oxy]-4H-chromen-4-one Formula: C₂₇H₃₀O₁₆</p>

RESULTS AND DISCUSSION

Spectrophotometric studies of DPPH-AS interaction

The decrease in absorbance of DPPH in the presence of antioxidant standards can be used for the calculation of the binding constant and the binding free energy, whereas the shift in wavelength values can be exploited for the determination of the mode of interaction, these observations are inspired from the study of the binding of drug molecules to DNA [11 – 14].

The interaction of AS with DPPH was studied by UV-Vis absorption titration for getting further clues about the mode of interaction and binding strength. The effect of different concentration of AS on the electronic absorption spectrum of a 10⁻⁴ M solution of DPPH in acetonitrile is shown in Figures 1, 2, 3, and 4. It was observed that DPPH had one strong absorption peak at 518 nm. After interaction with increasing amount of antioxidant standards, this peak decreased gradually and the wavelength had no obvious shift. The concentrations of antioxidants standards were chosen based on the optimal response in the electronic absorption spectrum.

Ascorbic acid-DPPH interaction

Figure 1 shows the electronic spectra of DPPH interaction with different concentrations of ascorbic acid.

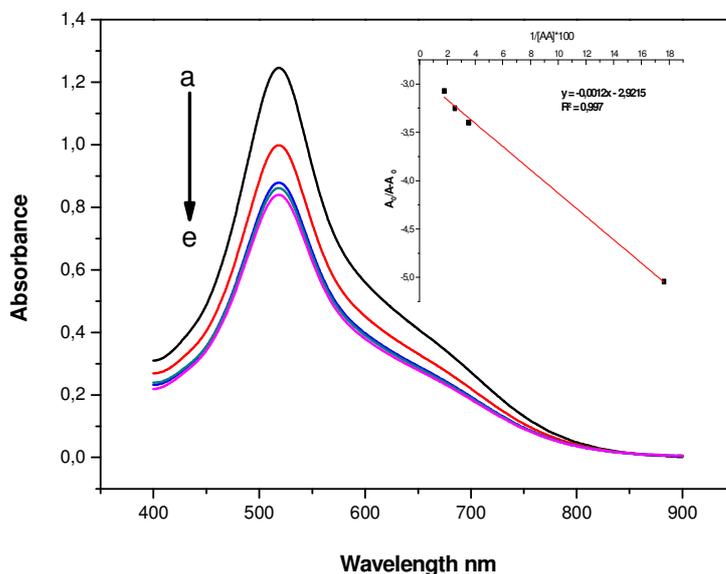


Figure 1. Electronic absorption spectra of 10^{-4} M DPPH interaction with 0 (a), 5.6×10^{-4} M (b), 2.8×10^{-3} M (c), 3.9×10^{-3} M (d), 5.6×10^{-2} M (e), AA in ACN, Inset: plot of $1/[AA]$ versus $A/A_0 - A$

Gallic acid-DPPH interaction

Figure 2 shows the UV spectra of DPPH interaction with different concentrations of gallic acid.

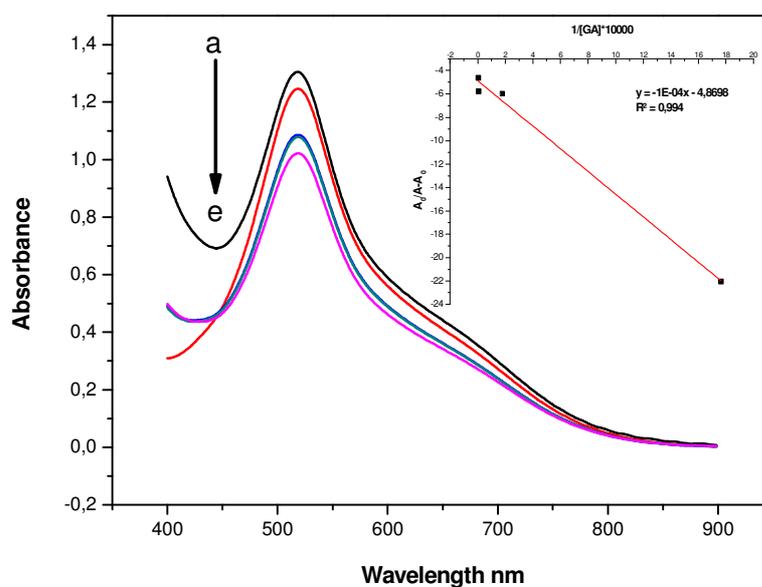


Figure 2. Electronic absorption spectra of 10^{-4} M DPPH interaction with 0 (a), 5.6×10^{-8} M (b), 2.8×10^{-6} M (c), 2.8×10^{-3} M (d), 4.5×10^{-3} M (e), GA in ACN, Inset: plot of $1/[GA]$ versus $A/A_0 - A$

Caffeic acid-DPPH interaction

Figure 3 shows the UV spectra of DPPH interaction with different concentrations of caffeic acid.

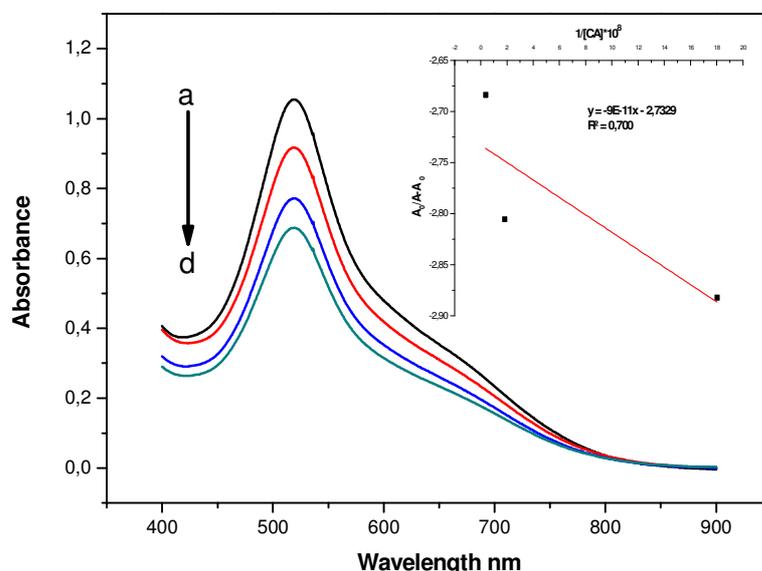


Figure 3. Electronic absorption spectra of 10^{-4} M DPPH interaction with 0 (a), 5.5×10^{-7} M (b), 5.5×10^{-6} M (c), 2.8×10^{-2} M (d), CA in ACN, Inset: plot of $1/[CA]$ versus $A/A_0 - A$

Figure 4 shows the UV spectra of DPPH interaction with different concentrations of rutin.

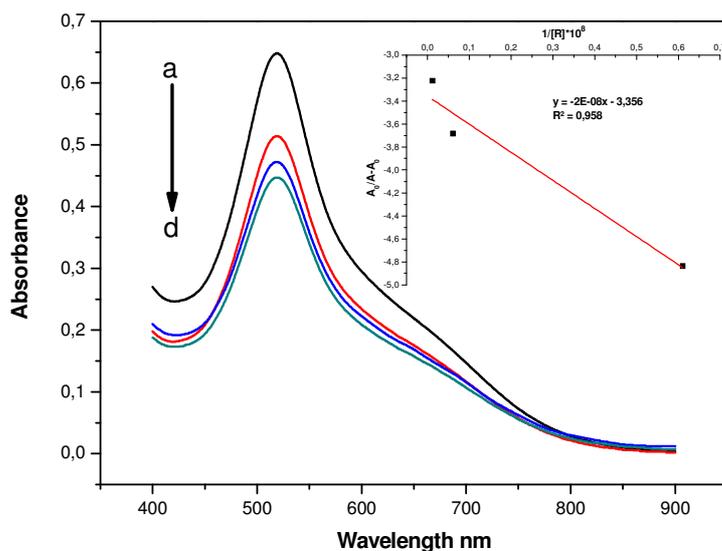


Figure 4. Electronic absorption spectra of 10^{-4} M DPPH interaction with 0 (a), 1.6×10^{-5} M (b), 1.6×10^{-4} M (c), 8.1×10^{-4} M (d), R in ACN,

Inset: plot of $1/[R]$ versus $A/A_0 - A$

Binding constant

The addition of gradually increasing concentration of AS in acetonitrile to a solution of 10^{-4} M of DPPH in the same solvent causes a remarkable overall decrease in absorbance (Figures 1, 2, 3 and 4). Based upon this decrease in absorbance, the binding constant was calculated according to Benesi-Hildebrand equation (1) used for the determination of the intrinsic binding constant/association constant of anticancer drug with DNA [15].

$$\frac{A_0}{A - A_0} = \frac{\epsilon_0}{\epsilon - \epsilon_0} + \frac{\epsilon_0}{\epsilon - \epsilon_0} \frac{1}{K[AS]} \quad (1)$$

where K is the binding constant, A_0 and A are absorbance of DPPH in the absence and in the presence of antioxidant standards, ϵ_0 and ϵ are their absorption coefficients respectively, $[AS]$ concentration of antioxidant standard.

The slope to intercept ratio of the plot between $A_0/(A - A_0)$ versus $1/[AS]$ yielded the binding constants, results are regrouped in Table 2. The moderate binding constants of ascorbic and gallic acids are indicative of electrostatic interaction, however higher values of binding constants of caffeic acid and Rutin are indicative of chemical interaction. The negative values of the Gibbs energy change signify the spontaneity of DPPH-AS interaction.

Table 2. Binding constants and binding free energy values

Compound	Equation	R ²	K [L·mol ⁻¹]	ΔG [KJ·mol ⁻¹]
DPPH-AA	$y = -1.2 \times 10^{-3}x - 2.9215$	0.997	24.3×10^2	-19.29
DPPH-GA	$y = -1.0 \times 10^{-4}x - 4.8698$	0.994	48.7×10^3	-26.70
DPPH-CA	$y = -9.0 \times 10^{-11}x - 2.7329$	0.700	30.4×10^9	-59.70
DPPH-R	$y = -2.0 \times 10^{-8}x - 3.3560$	0.958	16.8×10^7	-46.84

CONCLUSION

The binding parameters like binding constant and binding free energy of four commonly used antioxidant standards with 1,1-diphenyl-2-picrylhydrazyl (DPPH), were successfully determined using spectrophotometric based assay. The results indicated electrostatic interaction of DPPH radical with antioxidant standard ascorbic and gallic acid as the dominant mode, whereas caffeic acid and rutin interact chemically with DPPH.

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REFERENCES

1. Antolovich, M., Prenzler, P.D., Patsalides, E., McDonald, S., Robards, K.: Methods for Testing Antioxidant Activity, *Analyst*, **2002**, 127, 183-198;
2. Boubekri, C., Rebiai, A., Lanez, T.: Study of Antioxidant Capacity of Different Parts of two South Algerian Eggplant Cultivars, *Journal of Fundamental and Applied Sciences*, **2012**, 4 (2), 164-174;
3. Brand-Williams, W., Cuvelier, M.E., Berset C.: Use of a Free Radical Method to Evaluate Antioxidant Activity, *LWT - Food Science and Technology*, **1995**, 28 (1), 25-30;
4. Khelef, A., Lanez, T.: In vitro Assays of the Antioxidant Activities of Ferrocene Derivatives bearing Amine, Amide or Hydrazine Groups, *Der Pharma Chemica*, **2015**, 7 (6), 318-323;
5. Rebiai, A., Lanez, T.: Chemical Composition and Antioxidant Activity of *Apis Mellifera* Bee Pollen from Northwest Algeria, *Journal of Fundamental and Applied Sciences*, **2012**, 4 (2), 155-163;
6. Noda, Y., Anzai, K., Marl, A., Kohno, M., Shinmei, M., Packer, L.: Hydroxyl and Superoxide Anion Radical Scavenging Activities of Natural Source Antioxidants using the Computerized JES-FR30 ESR Spectrometer System, *Biochemistry and Molecular Biology International*, **1997**, 42 (1), 35-44;
7. Magnani, L., Gaydou, E.M., Hubaud, J.C.: Spectrophotometric Measurement of Antioxidant Properties of Flavones and Flavonols against Superoxide Anion, *Analytica Chimica Acta*, **2000**, 411 (1-2), 209-216;
8. Sánchez, J.C., García, R.F., Cors, M.T.: 1,1-Diphenyl-2-picrylhydrazyl Radical and Superoxide Anion Scavenging Activity of *Rhizophora mangle* (L.) bark, *Pharmacognosy Research*, **2010**, 2 (5), 279-84;
9. Shareef, M.I., Reddy P.J.M., Gopinath, S.M., Dayananda, K.S., Somashekhar, R., Mandal, A., Purushotham, K.M.: Superoxide Anion Scavenging Activity of *Carthamustinctorius* Flower, *International Journal of Innovative Research in Science, Engineering and Technology*, **2014**, 3 (3), 10101-10104;
10. Le Bourvellec, C., Hauchard, D., Darchen, A., Burgot, J.L., Abasq, M.L.: Validation of a New Method using the Reactivity of Electrogenerated Superoxide radical in the Antioxidant Capacity Determination of Flavonoids, *Talanta*, **2008**, 75, 1098-1103;
11. Lu, X., Zhu, K., Zhang, M., Liu, H., Kang, J.: Voltammetric Studies of the Interaction of Transition-metal Complexes with DNA, *Journal of Biochemical and Biophysical Methods*, **2002**, 52, 189-200;
12. Aslanoglu, M., Ayne, G.: Voltammetric Studies of the Interaction of Quinacrine with DNA, *Analytical and Bioanalytical Chemistry*, **2004**, 380, 658-663;
13. Palecek, E.: Past, present and future of nucleic acids electrochemistry, *Talanta - The International Journal of Pure and Applied Analytical Chemistry*, **2002**, 56, 809-819;
14. Ozsoz, M., Erdem, A., Kara, P., Kerman, K., Ozkan D.: Electrochemical Biosensor for the Detection of Interaction between Arsenic Trioxide and DNA based on Quanine Signal, *Electroanalysis*, **2003**, 15, 613-619;
15. Benesi, H.A., Hildebrand, J.H.: Spectrophotometric Investigation of the Interaction of Iodine with Aromatic Hydrocarbons, *Journal of the American Chemical Society*, **1949**, 71, 2703-2707.